Oxidative DNA Base Damage in Lymphocytes of HIV-infected Drug Users

PAWEL JARUGA^{a,*}, BARBARA JARUGA^b, ANITA OLCZAK^b, WALDEMAR HALOTA^b and RYSZARD OLINSKI^a

^aDepartment of Clinical Biochemistry, The Ludwik Rydygier Medical University, Karlowicza 24, 85-092 Bydgoszcz, Poland; ^bClinic of Infectious Diseases, The Ludwik Rydygier Medical University, Floriana 12, 85-061 Bydgoszcz, Poland

Accepted by Prof. M. Dizdaroglu

(Received 20 October 1998; In revised form 26 February 1999)

In the present study, we have studied the level of oxidative DNA base damage in lymphocytes of HIVinfected intravenous drug users (IDUs) and a seronegative control group. Chromatin was isolated from the lymphocytes and then analyzed by gas chromatography/isotope-dilution mass spectrometry with selectedion monitoring (GC/IDMS-SIM). Significantly greater levels of four oxidatively modified DNA bases were observed in chromatin samples from the symptomatic HIV-infected patients than in those from the seronegative patients. These were 5-hydroxyuracil, 5-hydroxycytosine, 8-hydroxyadenine and 8-hydroxyguanine. In the case of 5-hydroxyuracil and 8-hydroxyguanine, a statistically significant difference was also found between the control group and the asymptomatic HIVpositive patients. These results suggest that oxidative stress may play an important role in the pathogenesis of acquired immune deficiency syndrome (AIDS), and that administration of antioxidant drugs to HIVinfected patients may offer protection against AIDSrelated carcinogenesis.

Keywords: DNA base damage, gas chromatography/mass spectrometry, oxidative stress, reactive oxygen species, HIV/AIDS

INTRODUCTION

It is believed that patients infected with the human immunodeficiency virus (HIV) are under chronic oxidative stress.^[1] Oxidative stress is defined as a condition characterized by an increased production of reactive oxygen species (ROS), which may include the highly reactive hydroxyl radical (*OH) and hydrogen peroxide.^[2,3] ROS may be an important factor in mutagenesis and carcinogenesis.^[2,4] Moreover, there is evidence that oxidative stress mediated by ROS is one of the causes of apoptosis.^[5] ROS can cause DNA damage by modification of DNA bases.^[6] Some of the modified DNA bases have been found to possess premutagenic properties.^[7,8] Therefore, if they are not repaired within an appropriate time frame, they may contribute to carcinogenesis.^[9] Alternatively these modified bases may trigger apoptosis of the cell. Recently, it was proposed that induction of apoptosis

^{*} Corresponding author. Tel.: +48-52-341-4916. Fax: +48-52-341-5933. E-mail: pjaruga@aci.amb.bydgoszcz.pl.

might play a central role in pathogenesis of acquired immune deficiency syndrome (AIDS).^[10] It is well known that, when cellular DNA damage is not repaired, cells may be eliminated via apoptosis.^[1,11,12]

The direct cause of apoptotic death of lymphocytes in HIV infected patients is not well known. Intravenous drug users (IDUs) are one of the groups most endangered by HIV-infection. In this study, we have studied the level of oxidative DNA base damage in lymphocytes of HIV-infected, symptomatic and asymptomatic IDUs with that of seronegative IDUs (control group). The aim was to see whether oxidative DNA base damage is increased in HIV-infected patients when compared with control HIV-free individuals.

MATERIALS AND METHODS

Blood samples were obtained from a control group of nine male HIV-seronegative IDUs (mean age 27 years, range 18-36 years) and from seventeen male HIV-infected IDUs (mean age 30 years, range 23-35 years). All of the patients were hospitalized in the Provincial Hospital of Infectious Diseases. They were on a stable diet for the weeks before and during this investigation. Both groups of the patients used heroin during approximately the same time period. HIV-infected patients (HIV+) were classified according to the Centers for Disease Control surveillance definition^[13] [group IIa, (asymptomatic patients, n = 9), and group IIb (symptomatic patients n = 8)]. The mean CD4+ cell counts in asymptomatic patients and in symptomatic patients were 527/mm³ (range 364-861/mm³) and 146/mm³ (range 91-247/mm³), respectively. None of the patients had malignancies or signs of any infection. Chromatin was isolated from the lymphocytes and then analyzed by gas chromatography/isotope-dilution mass spectroscopy with selected-ion monitoring (GC/IDMS-SIM) as described.^[14,15]

RESULTS AND DISCUSSION

Using GC/IDMS-SIM technique, the following modified DNA bases were identified and quantified in chromatin samples isolated from lymphocytes: 5-hydroxyuracil (5-OH-Ura), 5-hydroxycytosine (5-OH-Cyt), 8-hydroxyadenine (8-OH-Ade) and 8-hydroxyguanine (8-OH-Gua). Considerable individual differences were observed in the levels of modified bases found in chromatin isolated from lymphocytes of the control group as well as the HIV-infected patients (Figure 1). Similar variations in the modified base levels were observed by us^[16-18] and others^[19] in DNA isolated from lymphocytes of cancerous and non-cancerous tissues of patients with different types of cancer. The levels of four modified DNA bases in the chromatin samples isolated from lymphocytes of the symptomatic patients were found to be significantly greater than those in chromatin samples from control patients (Figure 1). Table I shows the mean values and standard deviations calculated from the values in Figure 1. The levels of 5-OH-Ura and 8-OH-Gua were also significantly greater in asymptomatic HIV-infected patients than those in control patients (Figure 1, Table I). The greater levels of modified DNA bases may be a result of the reaction of endogenously produced ROS with cellular DNA.^[2] Since there is no evidence that AIDS is associated with a reduced ability to repair DNA damage, the observed increase could be a result of higher production of ROS in lymphocytes of HIVinfected patients. A wide variety of evidence supports the notion that oxidative stress is involved in the progression of AIDS. An excess of hydrogen peroxide combined with a deficiency in antioxidant enzymes, catalase and glutathione peroxidase, may lead to the overproduction of •OH,^[1] which in turn may be responsible for oxidative DNA damage.^[20] Typical products of reactions of *OH with DNA bases were detected in this work. These modified bases may possess premutagenic properties.^[7,8] Their increased production in lymphocytes of HIV-infected patients

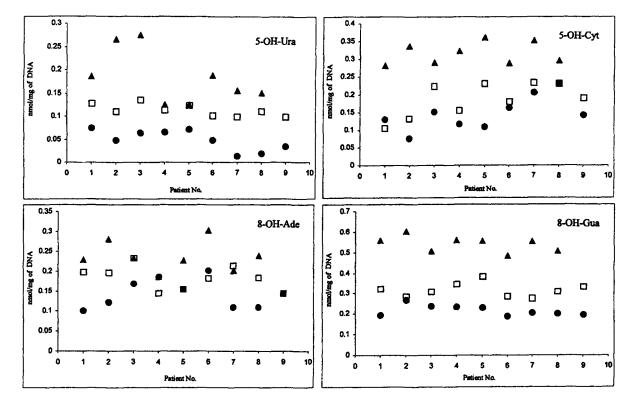


FIGURE 1 Levels of four modified DNA bases in lymphocytes of individual HIV negative (HIV-) (\oplus), positive IDUs: HIV+ (IIa) (asymptomatic patients) (\square), and HIV+ (IIb) (symptomatic patients) (\blacktriangle). The same individual has the same number in each plot and the numbers were given arbitrarily. One nmol of a lesion/mg of DNA corresponds to approximately 32 lesions/10⁵ DNA bases.

TABLE I Levels of modified DNA bases (nmol/mg of DNA) in lymphocytes of HIV negative (HIV–) and positive IDUs [HIV+ (IIa) (asymptomatic patients) and HIV+1 (IIb) (symptomatic patients)]

Modified base	HIV-	HIV+ (IIa)	HIV+ (IIb)
5-OH-Ura ^{a,b,c} 5-OH-Cyt ^{b,c} 8-OH-Ade ^{b,c} 8-OH-Gua ^{a,b,c}	$\begin{array}{c} 0.048 \pm 0.022 \\ 0.147 \pm 0.048 \\ 0.144 \pm 0.036 \\ 0.217 \pm 0.028 \end{array}$	$\begin{array}{c} 0.113 \pm 0.013 \\ 0.187 \pm 0.048 \\ 0.184 \pm 0.031 \\ 0.316 \pm 0.034 \end{array}$	$\begin{array}{c} 0.184 \pm 0.040 \\ 0.316 \pm 0.031 \\ 0.238 \pm 0.038 \\ 0.541 \pm 0.040 \end{array}$

The values represent the mean \pm standard deviation and were calculated from the values in Figure 1. One nmol/mg of DNA corresponds to 32 lesions/10⁵ DNA bases. The data were analyzed by the Student *t*-test (p < 0.05)

significant difference between HIV- and HIV+ (IIa);

^bsignificant difference between HIV+ (IIa) and HIV+ (IIb);

^csignificant difference between HIV- and HIV+ (IIb).

may be related to malignancies associated with AIDS, as HIV-infection predisposes an individual to several neoplasias, especially non-Hodgkin's lymphoma of B-cell origin.^[21-23] Recently, in-

creased levels of oxidatively modified DNA bases were found in cancerous and precancerous tissues, suggesting the involvement of oxidative DNA damage in cancer development.^[16,17]

It has been proposed that apoptosis initiated by oxidative stress is the direct cause of lymphocyte loss in patients infected with HIV.^[10,24] However, the mechanisms, which trigger apoptosis, have not been elucidated. One of the mechanisms for apoptotic death of cells may be unrepaired DNA damage. There is evidence that oxidative DNA damage induces apoptosis in murine T-cell hybridoma and in lymphocytes.^[25,26] Oxidatively modified DNA bases, which were found to have greater levels in HIV-infected patients than in control patients, may contribute to apoptosis. In this context, a recent study demonstrated that lymphocytes of HIV-infected individuals even at very early stages of infection were more susceptible to oxidative stress-mediated apoptosis than lymphocytes isolated from a control group.^[1]

In conclusion, significantly greater levels of oxidatively modified DNA bases were found in symptomatic HIV-infected IDUs than in seronegative IDUs. We postulate that the observed greater levels of oxidative DNA base damage may be one of the variables responsible for the apoptotic death of lymphocytes and carcinogenesis related to HIV-infection. This is the first study of oxidative DNA base damage in lymphocytes isolated from HIV-infected patients. The results support the notion that oxidative stress may play an important role in AIDS pathogenesis and the use of antioxidant drugs in the therapy of HIV-infected patients may offer protection against AIDS-related carcinogenesis and apoptosis.

Acknowledgements

This study was supported by grants from the Polish State Committee for Scientific Research (KBN), grant No. 4 P05A 090 14 and The Ludwik Rydygier Medical University in Bydgoszcz, Poland grant No. 59/97.

References

- G.W. Pace and C.D. Leaf (1995) The role of oxidative stress in HIV disease. Free Radicals in Biology and Medicine, 19, 523-528.
- [2] D.I. Feig, T.M. Reid and L.A. Loeb (1994) Reactive oxygen species in tumorigenesis. *Cancer Research*, 54, 1890–1894.
- [3] D.I. Feig, L.C. Sowers and L.A. Loeb (1994) Reverse chemical mutagenesis: identification of the mutagenic lesions resulting from reactive oxygen species-mediated damage to DNA. Proceedings of the National Academy of Sciences USA, 91, 6609–6613.
- [4] P.A. Amstad, G. Krupitza and P.A. Cerutti (1992) Mechanism of c-fos induction by active oxygen. *Cancer Research*, 52, 3952–3960.
- [5] D.M. Hockenbery, Z.N. Oltval, X. Yin, C.L. Milliman and S.J. Kosmeyer (1993) Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell*, 75, 241–251.
- [6] B. Halliwell and J.M.C. Gutteridge (1989) Free Radicals in Biology and Medicine, 2nd edn., Clarendon Press, Oxford.
- [7] S.S. Wallace (1998) Enzymatic processing of radiationinduced free radical damage in DNA. *Radiation Research*, 150, S60–S79.

- [8] D. Wang, D.A. Kreutzer and J.M. Essigman (1998) Mutagenicity and repair of oxidative DNA damage: insight from studies using defined lesions. *Mutation Research*, 400, 99–115.
- [9] R.A. Floyd (1990) The role of 8-oxoguanine in carcinogenesis. Carcinogenesis, 11, 1447–1450.
- [10] J.C. Ameisen and A. Capron (1991) Cell dysfunction and depletion in AIDS: The programmed cell death hypothesis. *Immunology Today*, 12, 102–105.
- [11] S.W. Lowe, E.M. Schmitt, S.W. Smith, B.A. Osborne and T. Jacks (1993) p53 is required for radiation-induced apoptosis in mouse thymocytes. *Nature*, 362, 847–849.
- [12] A.R. Clarke, C.A. Purdie, D.J. Harrison, R.G. Morris, C.C. Bird, M.L. Hooper and A.H. Wyllie (1993) Thymocyte apoptosis induced by p-53-dependent and independent pathways. *Nature*, 362, 849–852.
- [13] CDC (1993) Classification system for HIV infection and expended surveillance case definition for AIDS between adolescence and adults. *Morbidity and Mortality Weekly Report*, 41, 1–29.
- [14] E. Gajewski, G. Rao, Z. Nackerdien and M. Dizdaroglu (1990) Modification of DNA bases in mammalian chromatin by radiation-generated free radicals. *Biochemistry*, 29, 7876–7882.
- [15] M. Dizdaroglu (1994) Chemical determination of oxidative DNA bases damage by gas chromatography/mass spectrometry. *Methods in Enzymology*, 234, 3–16.
- [16] P. Jaruga, T.H. Zastawny, J. Skokowski, M. Dizdaroglu and R. Olinski (1994) Oxidative DNA base damage and antioxidant enzyme activities in human lung cancer. *FEBS Letters*, 341, 59–64.
- [17] R. Olinski, T.H. Zastawny, J. Budzbon, J. Skokowski, W. Zegarski and M. Dizdaroglu (1992) DNA base modifications in chromatin of human cancerous tissues. *FEBS Letters*, 193, 193–198.
- [18] R. Olinski, P. Jaruga, M. Foksinski, K. Białkowski and J. Tujakowski (1997) Epirubicin induced oxidative DNA damage and evidence for its repair in lymphocytes of cancer patients undergoing chemotherapy. *Molecular Pharmacology*, 52, 882–885.
- [19] D.C. Malins and R. Haimanot (1991) Major alterations in the nucleotide structure of DNA in cancer of the female breast. *Cancer Research*, 51, 5430–5432.
- [20] M. Dizdaroglu (1993) Chemistry of free radical damage to DNA and nucleoproteins. In: DNA and Free Radicals. (Eds. B. Halliwell and O.I. Aruoma), Ellis Horwood Ltd., London and New York, pp. 19-34.
- [21] T.F. Schulz, C.H. Boshoff and R.A. Weiss (1996) HIV infection and neoplasia. *Lancet*, 348, 587-591.
- [22] V. Beral, T. Peterman, R. Berkelman and H. Jaffe (1991) AIDS-associated non-Hodgkin lymphoma. *Lancet*, 337, 805–809.
- [23] C.K.O. Williams and L.O. Kashala (1994) AIDS-associated cancers. In: AIDS in Africa (Ed. M. Essex), Raven Press Ltd., New York, pp. 325–371.
- [24] T.S. Dobmeyer, S. Findhammer, J.M. Dobmeyer, S.A. Klein, B. Raffel and D. Hoelzer (1997) Ex vivo induction of apoptosis in lymphocytes is mediated by oxidative stress: role of lymphocyte loss in HIV infection. Free Radicals in Biology and Medicine, 22, 775-785.
- [25] R.L. Warters (1992) Radiation-induced apoptosis in a murine T-cell hybridoma. *Cancer Research*, 52, 883–890.
- [26] A.F. Slatter, C.S.I. Nobel and S. Orrenius (1995) The role of intracellular oxidants in apoptosis. *Biochimica Bio*physica Acta, 1271, 59-62.

200